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Attention/Zu Händen von/A l'attention de:	Date/Datum/Date: April 15, 2010
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☉ Urgent/ Dringend/ Urgent	☉ Reply ASAP/ Rückantwort/ Réponse urgente	☐ Please comment/ Erledigung/ Commentaires attendus	☐ Please review/ Überprüfung/ A vérifier	☐ For your information/ Kenntnisnahme/ Copie pour information

Total pages, including cover sheet: 16
Anzahi der übermittelten Seiten inkl. Deckblatt
Nombre de pages (Page de garde incluse)

Comments/Anmerkungen/Commentaires

RE: 10/536,853

Dear Ms. Martinez

Please find attached my proposed declaration used for support of the notion that flash chromatography implies plastic and/or glass tubes to a person of ordinary skill in that art. I would like to discuss with you to confirm that you and Examiner Langel agree that it does just that. The response is due over the weekend, so if we could have a brief discussion either later today or tomorrow, that would be great.

Sincerely, Pehr Jansson
Reg. No. 35,759
512 750-3046

PATENT

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Application No.	:	**10/536,853**
Applicant	:	**AZNAR, Pascal**
Filing Date	:	2005-05-27
Confirmation No.	:	4377
Art Unit	:	4116
Examiner	:	MARTINEZ, Brittany M
Docket No.	:	103.001
Customer No.	:	41754

DECLARATION UNDER 37 C.F.R. SECTION 1.132

I, Domingo Sanchez, Ph. D., declare and say:

That I am a citizen of Spain and I reside at Lilla Madviksvägen 34, 448 96 Tollered, Sweden.

That I graduated in 1978 from Chalmers University of Technology/Gothenburg University located in Göteborg, Sweden with a Ph.D degree in Organic Chemistry

That since 1984 I have been working in the field of chromatography for approximately 25 years, including development and marketing of silica-based packing materials to be used in the purification of pharmaceuticals by High Performance Liquid Chromatography, HPLC.

I am the author of approximately 50 peer-reviewed scientific papers and hold 5 patents, in the fields of Organic Chemistry and Chromatography.

I have been employed by Akzo Nobel from 1984. My work at Akzo Nobel has been in the field of chromatography.

I am very knowledgeable in regard to the fields of Flash Chromatography, High Performance Liquid Chromatography (HPLC), and Solid Phase Extraction (SPE).

That I am familiar with the above-identified patent application Serial Number 10/536,853 and with the references cited by the Examiner in the Office Action of December 17, 2010.

That the following accurately describes certain aspects of the field of Flash Chromatography:

- That Flash Chromatography operates at very low driving pressures - up to a typical maximum pressure of 10 bars - in contrast to, HPLC columns which operate at very high pressures - up to and sometimes exceeding 400 bars.
- That Flash Chromatography is typically performed using columns constructed from plastic or glass tubes or syringes not designed for high pressures.
- Early Flash Chromatography columns were made of glass, for example, such as those described in W. Clark Still, Michael Kahn and Abhijit Mitra, "Rapid Chromatographic Technique for Preparative Separations with Moderate Resolution", J. Org. Chem. Vol. 43, No 14, 1978 (Exhibit A).
- Modern Flash Chromatography columns are typically made of plastic. That is described, for example, in Horowitz, Gail, "Undergraduate Separations Utilizing Flash Chromatography" Journal of Chemical Education, Vol. 77, No. 2, February 2000, p. 263 (Exhibit B), and in "What is Flash Chromatography?", Labhut.com, http://www.labhut.com/education/flash/index.php, accessed on April 13, 2010 (Exhibit C).
- Other examples of plastic flash chromatography columns may be found in Varian Inc. SI™ Replacement Cartridges, http://www.varianinc.com/cgi-bin/nav?products/consum/flash/flashdisc/si&cid=LLIIJMNPFK , accessed on April 13, 2010 (Exhibit D) and Thomsom

Instrument Company, E. Merck Silica Syringe Barrel Columns, http://www.htslabs.com/catalog/index.php?d=flash/syringebarrel/emerck, accessed on April 13, 2010 (Exhibit E).

- That the Varian and Thomson columns illustrated in Exhibits D and E, respectively, are constructed from plastic.
- That the term *Flash Chromatography column* conveys to a person skilled in the art of chromatography a device constructed using a plastic or glass tube.

That the following accurately describes the differences between SPE and Flash Chromatography:

- SPE is a method where substances dissolved in a solvent are adsorbed to a powder material, namely irregular silica particles of large, ~40-60µm, particle size packed in open cartridges.
- That the Supelco reference (Supelco, Bulletin 910, Guide to Solid Phase Extraction cited in the Office Action) discloses only one material that is raw silica, namely, LC-Si (Supelco, Page 2).
- That I have visited and studied, on or before April 13, 2010, the relevant pages of the online catalog presented on the web site of Sigma-Aldrich (Supelco is a subsidiary of Sigma-Aldrich) to review the specifications for the LC-Si material.
- That the following web site is an online catalog for the Sigma-Aldrich's LC-Si material for SPE: http://www.sigmaaldrich.com/analytical-chromatography/analytical-products.html?TablePage=9642791

- That each of the products listed on that webpage is an *irregular* shaped silica material or a product using *irregular* shaped silica.

That the undersigned declares further that all statements made herein of his own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patents issuing thereon;

Further declarant saith not.

Date:

_____

Domingo Sanchez, Ph. D.

Exhibit A

(11) Potassium ferricyanide has previously been used to convert *vic*-1,2-dicarboxylate groups to double bonds. See, for example, L. F. Fieser and M. J. Haddadin, *J. Am. Chem. Soc.*, **86**, 2392 (1964). The oxidative dicarboxylation of 1,2-dicarboxylic acids is, of course, a well-known process. See inter alia (a) C. A. Grob, M. Ohta, and A. Weiss, *Helv. Chim. Acta*, **41**, 1911 (1958); and (b) E. N. Cain, R. Vukov, and S. Masamune, *J. Chem. Soc. D*, 98 (1969).

Rapid Chromatographic Technique for Preparative Separations with Moderate Resolution

W. Clark Still,* Michael Kahn, and Abhijit Mitra

Department of Chemistry, Columbia University, New York, New York 10027

Received January 26, 1978

We wish to describe a simple absorption chromatography technique for the routine purification of organic compounds. Large scale preparative separations are traditionally carried out by tedious long column chromatography. Although the results are sometimes satisfactory, the technique is always time consuming and frequently gives poor recovery due to band tailing. These problems are especially acute when samples of greater than 1 or 2 g must be separated. In recent years several preparative systems have evolved which reduce separation times to 1–3 h and allow the resolution of components having $\Delta R_f \geq 0.05$ on analytical TLC. Of these, medium pressure chromatography[1] and short column chromatography[2] have been the most successful in our laboratory. We have recently developed a substantially faster technique for the routine purification of reaction products which we call flash chromatography. Although its resolution is only moderate ($\Delta R_f \geq 0.15$), the system is extremely inexpensive to set up and operate and allows separations of samples weighing 0.01–10.0 g[3] in 10–15 min.[4]

Flash chromatography is basically an air pressure driven hybrid of medium pressure and short column chromatography which has been optimized for particularly rapid separations. Optimization studies were carried out under a set of standard conditions[5] using samples of benzyl alcohol on a 20 mm × 5 in. column of silica gel 60 and monitoring the column output with a Tracor 970 ultraviolet detector. Resolution is measured in terms of the ratio of retention time (r) to peak width (w,$w/2$) (Figure 1), and the results are diagrammed in Figures 2–4 for variations in silica gel particle size, eluant flow rate, and sample size.

A number of interesting facts emerge from these data. First, we find that one of the most popular grades of silica gel 60, 70–230 mesh (63–200 μm), gives the poorest resolution of any gel studied under our standard conditions. Second, particle sizes less than 40 μm offer no improvement in resolution with our method of packing.[7] Column performance is quite sensitive to the rate of elution and is best with relatively high eluant flow rates. The solvent head above the adsorbent bed should drop 2.0 ± 0.1 in./min for optimum resolution with mixtures of ethyl acetate/petroleum ether (30–60 °C).[8] Finally, the peak width shows the expected increase with the sample size. Sample recovery was ≥95%.

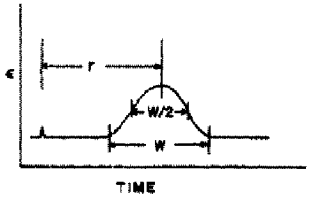

Figure 1. Typical chromatogram.

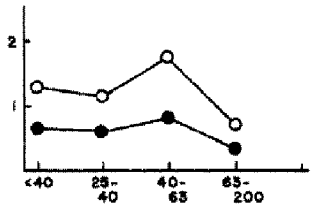

Figure 2. Silica gel particle size[6] (μm): (●) r/w; (○) $r/(w/2)$.

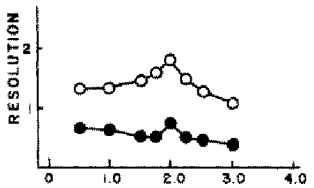

Figure 3. Eluant flow rate (in./min).

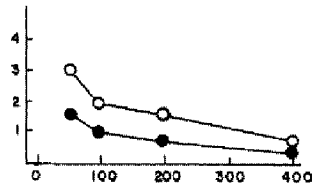

Figure 4. Sample size (mg).

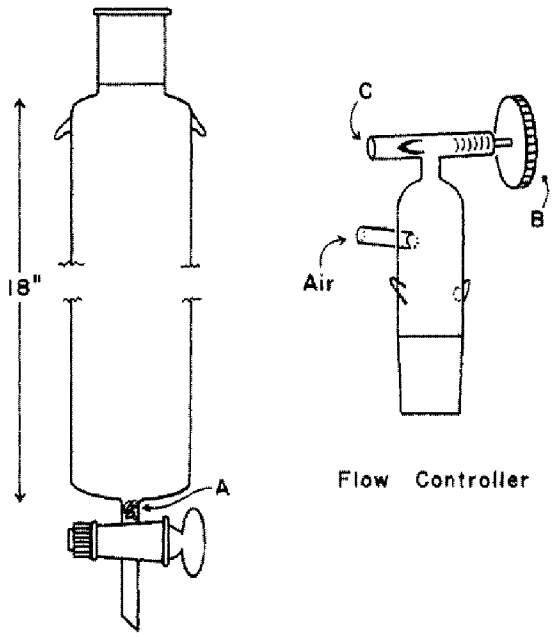

Figure 5.

The apparatus required for this technique consists of a set of chromatography columns and a flow controller valve (below). The column is a flattened bottom 18 in. glass tube fitted with a Teflon stopcock and topped with a 24/40 glass joint. Columns without fritted glass bed supports are generally preferred since they have significantly less dead volume than the standard fritted round-bottom variety. The flow controller

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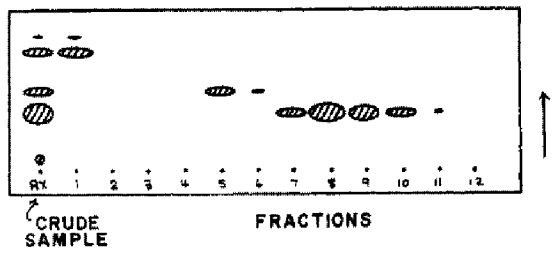


Figure 6.

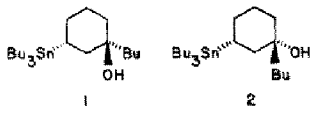
valve is a simple variable bleed device for precise regulation of the elution rate and is constructed from a glass/Teflon needle valve (Ace Glass Co. No. 8193-04 or equivalent) and a standard 24/40 joint.

A detailed procedure is presented in the experimental section and is summarized as follows: (1) A solvent is chosen which gives good separation and moves the desired component to R_f = 0.35 on analytical TLC (E. Merck No. 5765).[9] (2) A column of the appropriate diameter (see Table I) is selected and filled with 5–6 in. of dry 40–63 μm silica gel (E. Merck No. 9385).[10] (3) The column is filled with solvent and pressure is used to rapidly push all the air from the silica gel. (4) The sample is applied and the column is refilled with solvent and eluted at a flow rate of 2 in./min.

The time required to elute the desired components from the column is generally so fast (5–10 min) that we have abandoned automatic fraction collectors in favor of a simple rack holding forty 20 × 150 mm test tubes. Small fractions are typically collected early in the elution with larger ones being collected toward the end of the chromatography. Separated components are conveniently detected by spotting ~5 μL of each fraction along the long side of 7 cm × 2.5 cm TLC plate and then by developing the plate sideways. Heavier spotting may be required for small samples or highly retentive components. A typical separation is shown in Figure 6.

Over the past year we have run many hundreds of these columns. In every case we have been able to effect clean separation of compounds having $\Delta R_f \geq 0.15$ in less than 15 min and in many cases separations at $\Delta R_f \simeq 0.10$ were possible. The amount of sample used on a given column is proportional to its cross-sectional area and Table I can serve as a guide to column selection.

The sample size may increase substantially if less resolution is required; we have used a 50-mm column for the purification of up to 10 g of compound having impurities at $\Delta R_f \geq 0.4$. Resolution is maintained even with large diameter columns. For example the epimeric alcohols 1 and 2 have an R_f of 0.34



1 2

and 0.25, respectively, in 5% ethyl acetate/petroleum ether. A 1.0-g mixture of 1 and 2 (ΔR_f = 0.09) easily separated with only a 65-mg mixed fraction in 7 min on a 40-mm diameter column (500 mL of 5% EtOAc/petroleum ether).

If the components to be separated are closer on TLC than ΔR_f 0.15, increased resolution may be achieved by using a longer (e.g., 10 in.) column of gel alternatively a less polar solvent can be used. Such a solvent can be selected to move the desired components on TLC to R_f = 0.25 without increasing the elution times too drastically. In either case, the column should be only lightly loaded with sample and a rapid flow rate of 2 in./min should be maintained. Slower flows clearly give poorer resolution with ethyl acetate/petroleum ether mixtures.

Table I

column diameter, mm	vol of eluant,[a] mL	sample: typical loading (mg) $\Delta R_f \geq 0.2$	sample: typical loading (mg) $\Delta R_f \geq 0.1$	typical fraction size, mL
10	100	100	40	5
20	200	400	160	10
30	400	900	360	20
40	600	1600	600	30
50	1000	2500	1000	50

[a] Typical volume of eluant required for packing and elution.

In conclusion, flash chromatography provides a rapid and inexpensive general method for the preparative separation of mixtures requiring only moderate resolution. Even in cases where high resolution is required, preliminary purification by the flash technique allows simplified high-resolution separations without contamination of expensive HPLC columns. Finally, we would like to stress the facts that use of the 40–63 μm silica gel and a pressure- (and not vacuum-) driven flow rate of 2.0 in./min are crucial for successful separations by this method.

Experimental Section

Chromatography columns and the flow controller valve were assembled as described in the text. The silica gel used was 40–63 μm (400–230 mesh) silica gel 60 (E. Merck No. 9385).[10] Solvents were distilled prior to use. Thin layer chromatograms (TLC) were run on glass supported silica gel 60 plates (0.25-mm layer, F-254) (E. Merck No. 5765).

Flash Chromatography. General Procedure. First a low viscosity solvent system (e.g., ethyl acetate/30–60 °C petroleum ether)[8] is found which separates the mixture and moves the desired component on analytical TLC to an R_f of 0.35.[9] If several compounds are to be separated which run very close on TLC, adjust the solvent to put the midpoint between the components at R_f = 0.35. If the compounds are widely separated, adjust the R_f of the less mobile component to 0.35. Having chosen the solvent, a column of the appropriate diameter (see text, Table I) is selected and a small plug of glass wool is placed in the tube connecting the stopcock to the column body (A in the diagram above). Two telescoping lengths of glass tubing (6 and 8 mm o.d.) make placement of the glass wool plug easy. Next a smooth ⅛ in. layer of 50–100 mesh sand is added to cover the bottom of the column and dry 40–63 μm silica gel is poured into the column in a single portion to give a depth of 5.5–6 in. With the stopcock open, the column is gently tapped vertically on the bench top to pack the gel. Next a ⅛ in. layer of sand is carefully placed on the flat top of the dry gel bed and the column is clamped for pressure packing and elution. The solvent chosen above is then poured carefully over the sand to fill the column completely. The needle valve (B) of the flow controller is opened all the way and the flow controller is fitted tightly to the top of the column and secured with strong rubber bands. The main air line valve leading to the flow controller is opened slightly and a finger is placed fairly tightly over the bleed port (C). This will cause the pressure above the adsorbent bed to climb rapidly and compress the silica gel as solvent is rapidly forced through the column. It is important to maintain the pressure until *all* the air is expelled and the lower part of the column is cool; otherwise, the column will fragment and should be repacked unless the separation desired is a trivial one. Particular care is necessary with large diameter columns. The pressure is then released and excess eluant is forced out of the column above the adsorbent bed by partially blocking the bleed port (C). The top of the silica gel should not be allowed to run dry. Next the sample is applied by pipette as a 20–25% solution in the eluant to the top of the adsorbent bed and the flow controller is briefly placed on top of the column to push all of the sample into the silica gel.[11] The solvent used to pack the column is ordinarily reused to elute the column. The walls of the column are washed down with a few milliliters of fresh eluant, the washings are pushed into the gel as before, and the column is carefully filled with eluant so as not to disturb the adsorbent bed. The flow controller is finally secured to the column and adjusted to cause the surface of the solvent in the column to fall 2.0 in./min. This seems to be an optimum value of the flow rate for most low viscosity solvents for any column diameter with the 40–63 μm silica gel. Fractions are

collected until all the solvent has been used (see Table I to estimate the amount of solvent and fraction size). It is best not to let the column run dry since further elution is occasionally necessary. Purified components are identified as described in the text by TLC. If the foregoing instructions are followed *exactly*, there is little opportunity for the separation to fail.

Although we generally pack fresh columns for each separation, the expense of large-scale separations makes it advantageous to reuse large diameter columns. Column recycling is effected by first flushing (rate = 2 in./min) the column with approximately 5 in. of the more polar component in the eluant (generally ethyl acetate or acetone) and then with 5 in. of the desired eluant. If the eluant is relatively nonpolar (e.g., ≤10% EtOAc/petroleum ether), it may be more advisable to use a flushing solvent (e.g., 20–50% EtOAc/petroleum ether) which is somewhat less polar than the pure high polarity component.

Registry No.—1, 66417-28-5; 2, 66417-27-4.

References and Notes

(1) Such units have been described and used extensively by J. M. McCall, R. E. TenBrinkt, and C. H. Lin at the Upjohn Company and A. I. Meyers at Colorado State University.
(2) B. J. Hunt and W. Rigby, *Chem. Ind. (London)*, 1868 (1967).
(3) This is not a limitation but is merely the scale range which we have used.
(4) This is the total time required for column packing, sample application, and complete elution.
(5) Standard conditions: 5 in. high bed of 40–63 μm silica gel 60 in a 20 mm diameter column packed as described in text, 2.0 in. of solvent flow/min, 200 mg of benzyl alcohol, 25% ethyl acetate/petroleum ether eluant.
(6) These gels are manufactured by E. Merck and are the following grades: <40 μm (silica gel H, No. 7736), 25–40 μm (LiChroPrep Si60, No. 9390), 40–63 μm (silica gel 60, No. 9385), 63–200 μm (silica gel 60, No. 10180).
(7) Slurry packing, incremental dry packing, or single portion dry packing gave identical results with the 40–63 μm gel. Since the last technique was the simplest, it was employed in all our studies.
(8) This is a particularly good general solvent system. For extremely polar compounds, acetone/petroleum ether or acetone/methylene chloride mixtures are often useful. Significantly higher viscosity solvents will require slower optimum resolution flow rates.
(9) If this R_f is given by a solvent having <2% of the polar component, a slightly less polar eluant is desirable. Thus if 1% ethyl acetate/petroleum ether gives a compound an R_f of 0.35 on TLC, the column is run with 0.5% ethyl acetate.
(10) 40–63 μm gel is also used for medium pressure chromatography[1] and is available from MCB in 1 kg ($45/kg) or 25 kg ($16/kg) lots.
(11) If the sample is only partially soluble in the eluant, just enough of the more polar component is added to give complete dissolution. Large quantities of very polar impurities are best removed prior to chromatography so that excessive quantities of solvent or large increases in solvent polarity will be unnecessary for sample application.

Homo-*C*-nucleosides. The Synthesis of Certain 6-Substituted 4-Pyrimidinones[1]

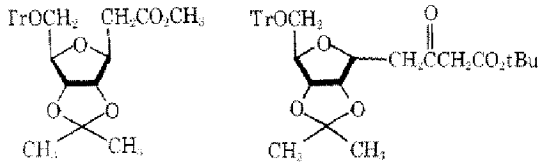
John A. Secrist III

Department of Chemistry, The Ohio State University, Columbus, Ohio 43210

Received February 1, 1978

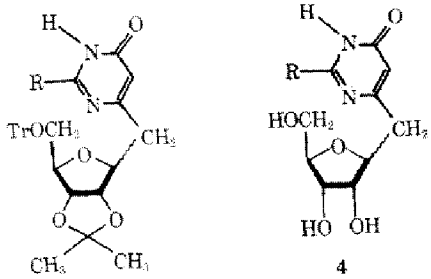
The chemistry of *C*-nucleosides has received considerable attention recently due to the biological activities of naturally occurring compounds such as showdomycin, formycin, and oxazinomycin.[2] Though synthetic methodology has evolved for the preparation of a number of *C*-nucleoside analogues,[2] only one investigation has dealt with the synthesis of homo-*C*-nucleosides,[3] compounds with a methylene unit between a carbon of the nitrogen base and the standard D-ribose moiety. This note describes the facile synthesis of a series of 6-substituted 4-pyrimidinone homo-*C*-nucleosides from the ester **1**, which is available in three steps from D-ribose.[4,5]

Treatment of **1** with lithio-*tert*-butyl acetate[6] in toluene at 0 °C for several hours affords an anomeric mixture (ca. 3:1, β/α) of the β-keto ester **2** in 75% yield. The assignment of β to the major anomer was made on the basis of ^{13}C NMR data. In particular, the isopropylidene methyls of the major anomer



1 2

Tr = trityl



3 4

a, R = NH_2
b, R = CH_3
c, R = SH
d, R = phenyl
e, R = H

occur at δ 25.66 and 27.54, within the range strongly indicative of a β configuration (25.5 ± 0.2 and 27.5 ± 0.2).[7,8]

It has been shown that the α-anomer of **1** is more stable than the β,[4] and recently a rationalization for this seemingly unusual behavior has been presented.[9] On this basis it seems likely that the α anomer of **2** is also more stable than the β. The conditions involved in the preparation of **2** (low-temperature, aprotic solvent) probably do not allow equilibration, though there is some leakage to the α-anomer. Further support for these postulates is provided by the finding that β-**2** is isomerized readily under basic conditions to an α/β mixture which is predominantly α.

Condensation of **2** with guanidine, acetamidine, thiourea, and benzamidine under basic conditions afforded the protected nucleosides **3a–d** as anomeric mixtures (ca. 3:1, α/β) which were chromatographically inseparable. That the major anomers after condensation are all α is also indicated by the chemical shifts of the isopropylidene methyls. For example, the shifts of the methyls in **3a** are at δ 25.09 and 26.33, clearly in the α range (24.9 ± 0.3 and 26.3 ± 0.2).[7,8] In view of the ready isomerization of β-**2** to a mixture of anomers containing predominantly α-**2**, it seems likely that equilibration is occurring prior to cyclization, and that the anomeric composition of **2** after equilibration dictates the ratio of α- and β-homo-*C*-nucleosides. Desulfurization of **3c** with Raney Nickel in refluxing 95% ethanol provided the hydrogen-substituted compound **3e**. Interestingly, while both urea and formamidine reacted with **2**, neither led to the formation of cyclized material under a variety of conditions. The free nucleosides **4a–e** were obtained by treatment of **3a–e** with either methanolic hydrogen chloride or aqueous trifluoroacetic acid for several hours. These acidic conditions, even over longer periods of time (2 days), caused no change in the α/β ratio of the nucleosides. Chromatographic separation of the free nucleoside anomers was once again not possible. **4e** was also available by desulfurization of **4c**.

The ^{13}C NMR spectra of the free nucleosides contained characteristic signals for the five compounds, and all values are reported in the Experimental Section. Salient ^1H NMR values are the methyl singlet of **4b** at δ 2.28 and the pyrimidine C_2H singlet of **4e** at δ 8.92, as well as the pyrimidine C_5 signal of all five nucleosides in the neighborhood of δ 6.0.

0022-3263/78/1943-2925$01.00/0

Undergraduate Separations Utilizing Flash Chromatography

Gail Horowitz

Department of Chemistry, Yeshiva University, New York, NY 10033; *gshorowit@ymail.yu.edu*

For a number of years, I have watched my organic chemistry laboratory students carry out gravity-based chromatographic separations and shared in their frustration as they watched their columns drip at a snail's pace. A simple separation of food dyes would often take one and a half to two hours to be completed. And while I felt it necessary that my students be familiar with this important technique, I found myself avoiding syntheses or procedures that required the use of column chromatography, both because I did not want to further tax the patience of my students and because of time constraints. I knew that flash chromatography apparatus was beyond the budget of my university and also worried about the safety of having undergraduates apply pressure to glass columns.

A few years ago, I happily discovered S. Todd Deal's article (*2*) discussing how to cheaply and easily implement the use of flash chromatography (*1*) in the undergraduate laboratory. I subsequently discovered similar articles (*3–8*) that had been published in this *Journal*. But having discovered an easy-to-use apparatus, it then became necessary to modify gravity-based chromatography experiments in order to carry them out using flash chromatography. Unfortunately, I did not find published experiments or articles giving procedures for undergraduate-level experiments involving flash chromatography. Having had to devise some such experiments on my own, I thought it would be helpful to other *Journal* readers to share my procedures with them.

Overview of Experiments

Two of the separations listed below involve natural products (spearmint and caraway, and spinach) and a third (benzil) is the starting step in the multistep syntheses of hexaphenylbenzene and dimethyl tetraphenylphthalate (*9, 10*). The carvone isolation exposes students to the subject of stereochemistry and how it relates to odor, and to the technique of polarimetry. The isolation of the components of spinach exposes students to the topics of chemistry of vision, photosynthesis, and UV–vis spectroscopy. The separation of β-carotene from tetraphenylcyclopentadienone (both colored components) vividly illustrates band separation to a beginning student witnessing column chromatography for the first time.

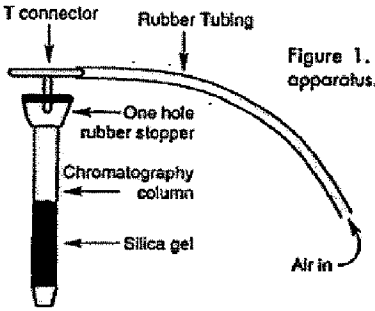


Figure 1. Flash chromatography apparatus.

In the procedures listed below, flash-grade silica gel[1] of mesh size 200–425 was used. The apparatus consisted of a polypropylene T connector,[2] one #6-sized one-hole rubber stopper, and a polypropylene 12 × 1.5-cm column (see Fig. 1).[3] (Use of polypropylene instead of glass is advantageous, in terms of both cost and safety.) Rubber tubing was used to connect the T connector to pressurized air and the air pressure was controlled by use of one's thumb. A flow rate of about 20 mL/min (2 in./min) was maintained.

Isolation of Carotenes, Chlorophylls and Xanthophylls from Spinach Extract

Two and a half grams of fresh spinach leaves were torn into small pieces and then stirred and crushed with a pestle in a mortar containing 10 mL of 95% ethanol. The ethanol was removed by vacuum filtration and the dehydrated spinach was added to 20 mL of hexane. The hexane solution was stirred and agitated for a few minutes and then the hexane was decanted from the spinach and concentrated to a volume of about 1 mL. The spinach extract was applied to a chromatography column packed with hexane and 5.5 g of silica gel. Three colored bands were eluted, corresponding to the carotenes (yellow), chlorophylls (bright green) and xanthophylls (yellow) (*11*). The solvents used were 90:10 hexane–EtOAc (ca. 30 mL), 70:30 hexane–EtOAc (ca. 50 mL), and 100% EtOAc (ca. 20 mL). The quality of separations varied depending on the care taken in column packing and sample loading. In some cases, it was possible to isolate an additional band (greenish gray) corresponding to the pheophytins, which eluted immediately after the carotenes. The colored bands collected were analyzed spectroscopically to determine yields. Yields on the order of 10–100 μg were obtained.

Separation of β-Carotene from Tetraphenylcyclopentadienone

Students were given a prepared mixture of β-carotene and tetraphenylcyclopentadienone dissolved in hexane.[4] Each student added 1 mL of this solution to a column packed with hexane and 7 g of silica gel. The β-carotene was eluted with approximately 50 mL of hexane. The solvent was then switched to 95:5 hexane–EtOAc and the tetraphenylcyclopentadienone fraction (about 15 mL) was collected.

Isolation of (+) and (–) Carvone from Caraway and Spearmint Oil (*12*)

Half a gram of caraway or spearmint oil[5] was dissolved in 1 mL of hexane and loaded onto a chromatography column packed with hexane and 6.5 g of silica gel. The column was eluted with hexane (approximately 30 mL) until limonene had ceased to elute (determined by TLC) and then carvone was eluted using ethyl acetate (approximately 25 mL). Students collected fractions in 13 × 100-mm test tubes. Optical purity of carvone product was determined by polarimetry.

Purification of Benzil from Benzoin

Starting with 1.4 mmol of benzoin, crude benzil was prepared by Cu^{2+}-catalyzed oxidation (*10*). The crude product was not very soluble in hexane and was added to the column as follows. The product was mixed with 1 g of flash-grade silica gel. Hexane was added until a slurry consistency was achieved. The beaker containing the hexane slurry was warmed with the palm of the hand and the slurry was stirred until the mixture had become homogeneous in color and the solvent had evaporated. The silica gel mixture was then poured onto the top of a column packed with hexane and 7 g of silica gel. Benzil was eluted using approximately 75 mL of 5% ethyl acetate in hexane.

Supplemental Material

Supplemental material for this article is available in this issue of *JCE Online*.

Notes

1. Fisher Scientific, cat. #S 733-1.
2. 5/16-in. o.d., Fisher Scientific, cat. #15-319 D.
3. Econo-Pac; Biorad, Hercules, CA, cat. #732-1010.
4. 0.040 g of β-carotene and 0.040 g of tetraphenylcyclopentadienone were dissolved in 100 mL of hexane.
5. Lorann Oils, Lansing, MI, cat. #2080, #0090.

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A Problem-Solving Approach to Chromatography in the Biochemistry Lab

W

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Many biochemistry lab manuals include experiments involving the purification of enzymes; wheat germ acid phosphatase (*1*, *2*) and lactic dehydrogenase (*3*, *4*) are common "targets". Most of these experiments rely mainly on precipitation techniques to effect purification and include, at most, a single chromatographic step. However, the purification of proteins in the "real world" usually involves a number of chromatographic steps and a number of chromatographic media (in addition to many other techniques). Thus, the typical experiment does not expose students to the idea of multiple-linked chromatographic steps in purification. These experiments also tend to be "cookbook" exercises in which the student follows a detailed protocol and does not have to directly address the "big picture".

The purification of enzymes is also a poor choice for introducing students to the correlation between the physical properties of molecules and their chromatographic behavior. Biological samples are complex mixtures of proteins, most of which are not visibly colored. Thus, students are unable to watch separations in real time and it is difficult to correlate the chromatographic behavior of a particular protein with its properties. At least one attempt to deal with these issues in the undergraduate biochemistry lab has been reported (*5*). However, this experiment requires the preparation of labeled proteins, a time-consuming step that many instructors might prefer to avoid.

Herein, I describe an experiment in which students must devise and carry out a separation protocol for a five-component mixture using multiple, sequential chromatographic steps. The components to be separated are well defined, commercially available, visibly colored compounds. Thus, students are able to watch the separation of the compounds and to correlate their movement on various chromatographic media.

Students are given mixtures of colored molecules and three chromatographic matrices (a cation exchange medium, an anion exchange medium, and a size exclusion medium). After an extensive prelab lecture dealing with the mechanics of chromatography, students are challenged to devise (and carry out) a procedure for the separation of the five colored molecules whose properties they are given (see Table 1).

Students are told to proceed in two stages over two 3-hour lab periods. For the first stage, each student is given 1-mL samples of two test mixtures, each of which contains three components (see Table 1). They are instructed to run a portion of each sample on each of the chromatographic matrices provided, to collect fractions, and to make careful observations of the separations.

What is Flash Chromatography?

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Simple Definition: Flash Chromatography is a rapid form of preparative column chromatography based on optimised pre-packed columns through which is pumped solvent at a high flow rate. It is a simple and economical approach to Preparative LC.

The technique was published in the Journal of Organic Chemistry in 1978 as an alternative to simple column chromatography. View abstract.

For purifying organic compounds, flash chromatography is a quick and inexpensive technique. Initially developed in 1978 by W.C. Stills of Columbia University, flash chromatography is now a popular method of purification and separation using normal phases. Increasingly, the use of reverse phase packing materials is opening up the technique to a wider range of preparative separations.

Flash chromatography utilises a plastic column filled with some form of solid support, usually silica gel, with the sample to be separated placed on top of this support. The rest of the column is filled with an isocratic or gradient solvent which, with the help of pressure, enables the sample to run through the column and become separated. Flash chromatography used air pressure initially, but today pumps are used to speed up the separation. This technique is considered a low to medium-pressure technique and may be scaled up for separations from a few mg to many tens or hundreds of grammes.

Applications are many and varied from drug discovery, sample clean-up, natural product purification and many more.

There is a direct relationship for normal phase flash chromatography with TLC and often the techniques are used together prior to and post flash separations.

The following pages are designed to offer an Introduction to Flash Chromatography.

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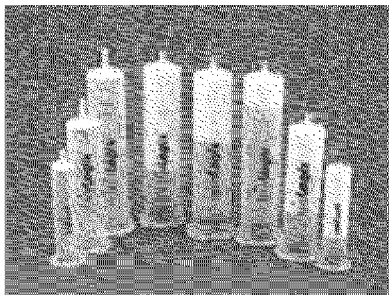

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